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## Brief Report

### Reactive oxygen species and cell morphology of MC3T3E1 pre-osteoblast cell line exposed to methylglyoxal by laser scanning confocal microscopy

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#### Key Words

AGEs precursor; MC3T3E1;  
Methylglyoxal; Pre-osteoblast

#### Abstract

Advanced glycation end products (AGEs) are known to increase in osteoporosis. Methylglyoxal is AGEs precursor that is toxic to osteoblasts. Methylglyoxal toxicity in osteoblasts involves oxidative stress. The involvement of reactive oxygen species (ROS) in changes of MC3T3E1 pre-osteoblast cell line morphology caused by methylglyoxal exposure is still unknown. This study was aimed to investigate a possible effect of methylglyoxal on ROS level and morphology of MC3T3E1 cell line. Pre-osteoblast MC3T3E1 cell line, obtained from American Type Culture Cell (ATCC), was exposed to methylglyoxal at several concentrations. Then ROS level and cell morphology were evaluated by laser scanning confocal microscopy using H<sub>2</sub>DCFDA. The optimal dose and exposition time of MC3T3E1 pre-osteoblast cell subclones to methylglyoxal is estimated to be 5 μM for 6 h which showed fusiform, polygonal cells, a homogenous cytoplasm and a clear hole as nucleus in the center of the cell. In conclusion, methylglyoxal exposure of MC3T3E1 pre-osteoblast cell line increased ROS level at 5 μM for 6 h of exposure. Beginning with this dose, the cell presents morphologic markers of apoptosis.

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## INTRODUCTION

Methylglyoxal is an electrophilic reactive substance which attacks functional groups of macromolecules to change their biological activity. Protein modification by methylglyoxal induces loss of enzymatic activity, change of conformational structure, and development of fluorescence [1]. Methylglyoxal modifies lysine and arginine residues leading to changes in their characteristic structure [2]. At cellular level, methylglyoxal induces the arrest of cell growth, dysfunction, apoptosis, necrosis, mechanisms that almost involve protein modification [3]. In addition, methylglyoxal can irreversible bind into protein amino acids and nucleic acids resulting in methylglyoxal-

derived advanced glycation end products (MAGEs) [4].

Oxidative stress due to reactive oxygen species (ROS) is a main factor for decreasing physiological functions such as ion transport, electrical activity and cellular signal transduction [5]. Previous *in vitro* studies showed that AGEs interaction with their receptors (RAGEs) induce oxidative stress and activation of the transcription factor NF-κB. Besides that, AGEs induces antioxidant depletion [6]. The effect of methylglyoxal on mature osteoblasts is mediated by apoptosis through biochemical pathways, including c-Jun N-terminal kinase (JNK) activity, change of mitochondria membrane, release of cytochrom C, increased Bax/Bcl-2 ratio, and the activation of caspase 3 and 9. A

previous *in vivo* study showed that administration of methylglyoxal at concentrations of 100-200  $\mu\text{g}$  via drinking water increases loss of bone mineral density of rats [7].

As far we know, there are no investigations on the effects of methylglyoxal exposure on ROS of MC3T3E1 pre-osteoblast cell line. To understand the involvement of methylglyoxal in osteoporosis, this study was aimed to evaluate the effect of methylglyoxal exposure on possible morphological changes and ROS of MC3T3E1 pre-osteoblast cell line by laser scanning confocal microscopy.

## MATERIAL AND METHODS

### Cell line

MC3T3E1 pre-osteoblast cell line subclone 4 was obtained from *American Type Culture Cell Collection* (ATCC). MC3T3E1 pre-osteoblast cell line was exposed to methylglyoxal at concentrations 2.5, 5, 10 and 20  $\mu\text{M}$  for 1, 3, 6 and 12 h of exposition time, according to previous studies [7, 8]. The control group was set as MC3T3E1 pre-osteoblast cell line subclone 4 without methylglyoxal exposure.

### Cell culture

MC3T3E1 pre-osteoblast cell line subclone 4 was grown in 75  $\text{cm}^2$  flasks in atmosphere humidification at 5%  $\text{CO}_2$  in Dulbecco's modified Eagle's medium (DMEM) supplemented by 100 U/ml penicilin, 100  $\mu\text{g}/\text{ml}$  streptomycin and 10% (v:v) fetal bovine serum (FBS). DMEM and FBS were obtained from Gibco® (Invitrogen). Subculture was done using 0.1% trypsin 1 mM EDTA in phosphate buffer saline free of  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ . In 70-80% confluent cultures, cell was harvesting into 6 or 24 well. These culture cell methods were done in Central Laboratory of Life Science, University of Brawijaya, Malang, Indonesia.

### Reactive oxygen species analysis

ROS analysis was done using confocal laser scanning microscope [9]. For this reason, 25  $\mu\text{M}$  2,7-dichlorodihydrofluorescein diacetate ( $\text{H}_2\text{DCFDA}$ ) was added to MC3T3E1 pre-osteoblast cell line subclone 4. After that, cells were washed with PBS and fixated by 10% buffer formalin for 10 min. Mounting was done on slide glass by ProLong® Antifade (Invitrogen). The evaluation of cell morphology was done using Zeiss LSM 510 laser scanning confocal microscopy. The green fluorescence in MC3T3E1 pre-osteoblast cell line subclone 4, expressed as arbitrary units, was estimated as ROS level.

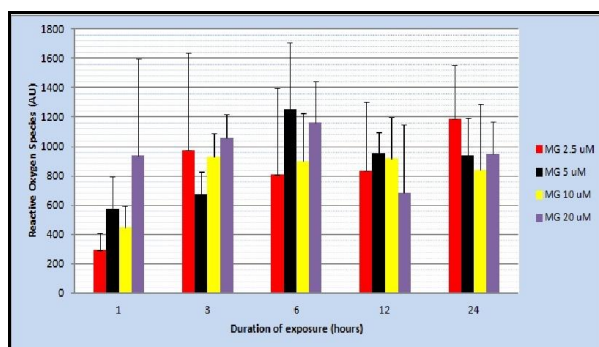
## RESULTS

### Reactive oxygen species level

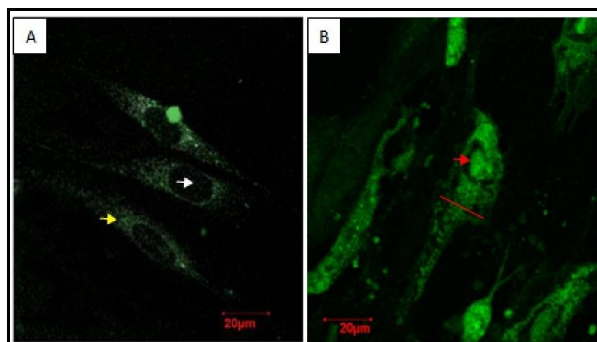
The highest level of ROS was achieved at 5  $\mu\text{M}$  for 6 h of exposure of the MC3T3E1 pre-osteoblast cell line subclone 4 to methylglyoxal (Fig.1). However, the levels of ROS is not significantly different between groups ( $P > 0.05$ ).

### Morphology of MC3T3E1 pre-osteoblast cell line subclone 4

Morphology of normal MC3T3E1 pre-osteoblast cell line subclone 4 was regular fusiform, polygonal; the surface of cytoplasm was homogenous and clear hole as nucleus was found in the center of the cells. At highest level of ROS, which achieved at 5  $\mu\text{M}$  for 6 h, the cell showed swelling, oval shape, and cytoplasm condensation as morphology markers for apoptosis (Fig.2).



**Figure 1.** The level of ROS in methylglyoxal-exposed cells; values are presented as mean  $\pm$  SD; MG: methylglyoxal; AU: arbitrary units. The optimal dose of methylglyoxal for highest production ROS in MC3T3E1 pre-osteoblast cell line subclone 4 is 5  $\mu\text{M}$  for 6 h of exposure.



**Figure 2.** Confocal micrograph of MC3T3E1 pre-osteoblast cell line subclone 4 not exposed (A) and exposed to methylglyoxal at optimal dose (B). Morphology of normal MC3T3E1 pre-osteoblast cell line subclone 4 was regular fusiform, polygonal; the surface of cytoplasm was homogenous (yellow arrow) and clear hole as nucleus was found in the center of cell (white arrow). At this optimal dose, the morphology of MC3T3E1 pre-osteoblast cell line subclone 4 showed swelling, oval shape (red line), and cytoplasm condensation as apoptosis marker (red arrow). [Magnification x400; bar is 20  $\mu\text{m}$ ]

## DISCUSSION

The main finding of this study is that methylglyoxal was able to increase ROS in MC3T3E1 pre-osteoblast cell line although not reached statistically difference. In addition, methylglyoxal also changed the morphology of MC3T3E1 pre-osteoblast cell line at the optimal dose. Increasing of ROS was probably due to activation of NADPH oxidase [6]. This activation resulted in superoxide radical catalyzed by its endogenous antioxidant superoxide dismutase forming hydrogen peroxide or another ROS. Besides, a previous study showed that the reaction between methylglyoxal and arginine increases methylglyoxal radical and superoxide radical [10]. The optimal dose of methylglyoxal exposure was found to be 5  $\mu$ M for 6 h of exposure. This finding indicates that ROS will achieve intracellular maximum levels at this dose. The levels of ROS was the result of the depletion of glutathione and the activity of a glyoxalase system for conversion and detoxification of methylglyoxal in cytoplasm [10-12]. Methylglyoxal was toxic to the cell due to induction of ROS for apoptosis signaling [13]. Former studies showed that methylglyoxal induced cytotoxicity at longer times of exposure (4-24 hours) [14, 15], whereas it was not toxic in short time of exposure to myoblast L6 cell line [13], hepatocytes [16], and human endothelial cells [17].

The normal morphology of MC3T3E1 pre-osteoblast cell line is fusiform, polygonal and regular shaped. The increase of ROS exposure induced swelling and apoptosis. A recent study reported that at 400  $\mu$ mol/l of H<sub>2</sub>O<sub>2</sub> exposure no marked morphological change was observed under a light microscope; cells started swelling with 800-1000  $\mu$ mol/l; and at 1000  $\mu$ mol/l, cells underwent apoptosis [18]. In the present study, the morphology of MC3T3E1 pre-osteoblast cell line at highest intracellularly ROS level from methylglyoxal exposure resulted in swelling, oval shape, cytoplasm condensation, and the margin between cytoplasm and nucleus became unclear as morphologic markers of apoptosis.

In conclusion, methylglyoxal exposure at MC3T3E1 pre-osteoblast cell line increased ROS level at 5  $\mu$ M for 6 h of exposure. At this dose, the cells start showing evidence of morphologic apoptosis markers.

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## CONFLICT OF INTEREST

None to declare

## REFERENCES

- Kalapos MP. The tandem of free radical and methylglyoxal. *Chem-Biol Interact* 2008; 171:251-71.
- Chan W, Wu H, Shiao N. Apoptotic signaling in methylglyoxal-treated human osteoblasts involves oxidative stress, c-jun N-terminal kinase, caspase-3, and p21-activated kinase 2. *J Cell Biochem* 2007; 100:1056-69.
- Schalkwijk CG, Bezu J, van der Schors RC, Uchida K, Stehouwer CDA, van Hinsbergh VWM. Heat-shock protein 27 is major methylglyoxal-modified protein in endothelial cells. *FEBS Lett* 2011; 580:1565-70.
- Oliveira LMA, Lages A, Gomes RA, Neves H, Familia C, Coelho AV, Quintas A. Insulin glycation by methylglyoxal results in native-like aggregation and inhibition of fibril formation. *BMC Biochem* 2011; 12:41.
- Nam SH, Jung SY, You C, Ahn EH, Suh CK. H<sub>2</sub>O<sub>2</sub> enhances Ca<sup>2+</sup> release from osteoblast internal stores. *Yonsei Med J* 2002; 43:229-35.
- Singh R, Barden A, Mori T, Beilin L. Advanced glycation end products: a review. *Diabetologia* 2001; 44:129-46.
- Chan W, Wu H. Protective effects of curcumin on methylglyoxal-induced oxidative DNA damage and cell injury in human mononuclear cells. *Acta Pharmacol Sinica* 2006; 27:1192-8.
- Mercer N, Ahmed H, Etcheverry SB, Vasta GR, Cortizo AM. Regulation of advanced glycation end product (AGE) receptors and apoptosis by AGEs in osteoblast like cells. *Mol Cell Biochem* 2007; 306:87-94.
- Kristiansen KA, Jensen PE, Moller IM, Schulz A. Monitoring reactive oxygen species formation and localisation in living cells by use of the fluorescent probe CM-H<sub>2</sub>DCFDA and confocal laser microscopy. *Physiol Plant* 2009; 136(4):369-83.
- Yim H, Kang S, Hah Y, Chock PB, Yim MB. Free radicals generated during the glycation reaction of amino acids by methylglyoxal. *J Biol Chem* 1995; 270:28228-33.
- Lee C, Yim MB, Chock PB, Yim HS, Kang SO. Oxidation reduction properties of methylglyoxal-modified protein in relation to free radical generation. *J Biol Chem* 1998; 273:25272-8.
- Oya T, Hattori N, Mizuno Y, Miyata S, Maeda S, Osawa T, Uchida K. Methylglyoxal modification of protein. Chemical and immunochemical characterization of methylglyoxal-arginine adducts. *J Biol Chem* 1999; 274:18492-502.
- Riboulet-Chavey A, Pierron A, Durand I, Murdaca J, Giudicelli J, Oberghen EV. Methylglyoxal impairs the insulin signaling pathways independently of the formation of intracellular reactive oxygen species. *Diabetes* 2006; 55:1289-99.
- Liu BF, Miyata S, Hirota Y, Higo S, Miyazaki H, Fukunaga M, Hamada Y, Ueyama S, Muramoto O, Uriuhara A, Kasuga M. Methylglyoxal induces apoptosis through activation of p38 mitogen-activated protein kinase in rat mesangial cells. *Kidney Int* 2003; 63:947-57.
- Fukunaga M, Miyata S, Liu BF, Miyazaki H, Hirota Y, Higo S, Hamada Y, Ueyama S, Kasuga M. 2004. Methylglyoxal induces apoptosis through activation of p38 MAPK in rat Schwann cells. *Biochem Biophys Res Comm* 2004; 320:689-95.
- Shangari N, O'Brien PJ. The cytotoxic mechanism of glyoxal involves oxidative stress. *Biochem Pharmacol* 2004; 68:1433-42.

17. Portero-Otin M, Pamplona R, Bellmunt MJ, Ruiz MC, Prat J, Salvayre R, Negre-Salvayre A. Advanced glycation end product precursor impair epidermal growth factor receptor signalling. *Diabetes* 2002; 51:1535-42.
18. Lin J, Fan Y, Mehl C, Zhu J, Chen H, Jin L, Xu J, Wang H. *Eucommia ulmoides* Oliv. Antagonizes H<sub>2</sub>O<sub>2</sub>-induced rat osteoblastic MC3T3-E1 apoptosis by inhibiting expressions of caspases 3, 6, 7, and 9. *J Zhejiang Univ-Sci B* 2011; 12:47-54.

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